

## CHO-K1 Human CCR8 Cell Line

CHO-K1 Human CCR8 Cell Line provides an engineered cellular model for investigating the receptor system specified in its construct.

GPCR signaling connects extracellular ligand recognition with intracellular responses.

Applications include receptor pharmacology, agonist or antagonist profiling, and development of cell-based functional assays.

|                     |                       |
|---------------------|-----------------------|
| CTDS SKU            | CTDS91364             |
| Pack size           | 1 x 1 million cells   |
| Product category    | Engineered Cell Lines |
| Product subcategory | GPCR Cell Lines       |
| Detailed category   | Class A (Rhodopsin)   |

### PRODUCT BACKGROUND

CHO-K1 Human CCR8 Cell Line is an engineered GPCR cell model expressing or functionally linked to CCR8 for controlled in vitro research. It is suitable for target and pathway characterization, cell-based screening, pharmacology, potency testing and assay development. Reporter, resistance or other engineered features stated in the product name can support quantitative or comparative functional readouts. For research use only.

### BIOLOGICAL CHARACTERISTICS

|                              |                                               |
|------------------------------|-----------------------------------------------|
| Host cell                    | CHO-K1                                        |
| Alternate name 1             | GPCR engineered Stable Cell Line              |
| Cell growth type             | Adherent Cells                                |
| Biosafety level              | BSL-2                                         |
| Tumorigenicity / cell status | Normal Cells                                  |
| Target / construct           | Stable human CCR8 target-expressing cell line |

### CULTURE REQUIREMENTS

|                 |                                                                                                                                                             |
|-----------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Culture medium  | Ham's F-12K (Kaighn's) + 10% FBS; for engineered/stable lines, maintain the product-specific CTDS selection antibiotic(s) at the recommended concentration. |
| Freezing medium | 90% FBS + 10% DMSO                                                                                                                                          |

### RECEIPT AND CRYOPRESERVATION

Upon Receipt of Cells

1 mL cryovials are packed in dry ice for transportation. After receipt, clean the cryovial with 70% IPA to disinfect. Stored in a -80°C refrigerator overnight and then transfer to liquid nitrogen or revive them directly as per your use.

Note If the dry ice is found to have evaporated or the cryovial cap is detached/ damaged or the cells seems damaged, please contact us immediately.

### CULTURE AND RECOVERY INSTRUCTIONS

Recovery of Frozen Cells:

For Suspension Cell passaging, the following methods can be used as reference:

- 1) Shake the cryotube containing 1mL of cell suspension quickly within seconds in a 37°C water bath to thaw.
- 2) Add it to a centrifuge tube containing 4-6mL of complete medium and mix well. Centrifuge at 1000rpm for 3-5mins.
- 3) Discard the supernatant, and resuspend the cells in complete medium.
- 4) Then add the cell suspension to a culture flask (or dish) containing 6-8ml of complete medium and culture overnight at 37°C.
- 5) Observe the cell growth and cell density under a microscope the next day.
- 6) Cell subculture: If the cell density reaches 80%-90%, subculture can be performed.

For Adherent Cell passaging, the following methods can be used as reference:

- 1) Shake the cryotube containing 1mL of cell suspension quickly within seconds in a 37°C water bath to thaw.
- 2) Add it to a centrifuge tube containing 4-6mL of complete medium and mix well. Centrifuge at 1000rpm for 3-5mins.
- 3) Discard the supernatant, and rinse the cells 1-2 times with PBS without calcium and magnesium ions.
- 4) Add 0.25% (w/v) trypsin-0.53 mM EDTA to the culture flask (1-2 mL for T25 flask, 2-3 mL for T75 flask),
- 5) Place in a 37°C incubator for digestion for 1-2 minutes (difficult-to-digest cells can be properly prolonged for digestion time),
- 6) Observe the cell digestion under a microscope. If most of the cells become round and fall off, quickly take it back to the operating table, tap the culture flask a few times, and add 3-4 ml of culture medium containing 10% FBS to terminate digestion.
- 7) Gently mix and aspirate, centrifuge at 1000rpm for 3-5min, discard the supernatant. Add 1-2mL culture medium and mix evenly.
- 8) Divide the cell suspension into new T25 bottles at a ratio of 1:2.
- 9) Add 6-8ml of new complete culture medium prepared according to the instructions to maintain cell growth vitality, and perform subsequent passages at a ratio of 1:2~1:5 as per actual conditions.

## CHARACTERIZATION AND QUALITY INFORMATION

|                              |                                                                                                                             |
|------------------------------|-----------------------------------------------------------------------------------------------------------------------------|
| Sterility                    | Bacteria: Negative                                                                                                          |
|                              | Yeast: Negative                                                                                                             |
|                              | Mycoplasma: Negative                                                                                                        |
| Pathogen testing             | HIV: Negative                                                                                                               |
|                              | Hepatitis B: Negative                                                                                                       |
| Markers / assay readout      | HTRF IP-One functional assay; Calcium-Mobilization Functional Assay; cAMP Functional Assay; Beta-Arrestin Recruitment Assay |
| Assay / validation method    | HTRF IP-One Functional assay; Calcium-Mobilization Functional assay; cAMP Functional assay; Beta-arrestin Recruitment Assay |
| Authentication / QC evidence | Sterility: Bacteria: Negative                                                                                               |
|                              | Yeast: Negative                                                                                                             |
|                              | Mycoplasma: Negative; Pathogen testing: HIV: Negative Hepatitis B: Negative                                                 |

## HANDLING AND STORAGE

|              |                                                          |
|--------------|----------------------------------------------------------|
| Shelf life   | 12 months at time of shipping                            |
| Shipping     | Shipped on Dry Ice at -20°C                              |
| Storage      | Store at -176°C.                                         |
| Intended use | For Research Use Only. Not for diagnostics or human use. |

## PRODUCT DISCLAIMER

The data indicated herein are as indicated and validated in our laboratory. As these cell lines are of human/animal origin, they carry a risk of latent human/animal viruses (e.g., Epstein-Barr virus, hepatitis B). Therefore they should be handled accordingly with precaution. These reagents are for research use only and not for in-vitro diagnostics or human use.

## ADDITIONAL CLASSIFICATION

|                  |                            |
|------------------|----------------------------|
| Product grouping | Engineered Cell Lines      |
| Product subgroup | GPCR   Class A (Rhodopsin) |

Not a lot-specific certificate of analysis.